

A Patient with a History of Pulmonary *Mycobacterium avium* Complex Infection and Chronic Fever

CIS 2016 PID Summer School Presentation

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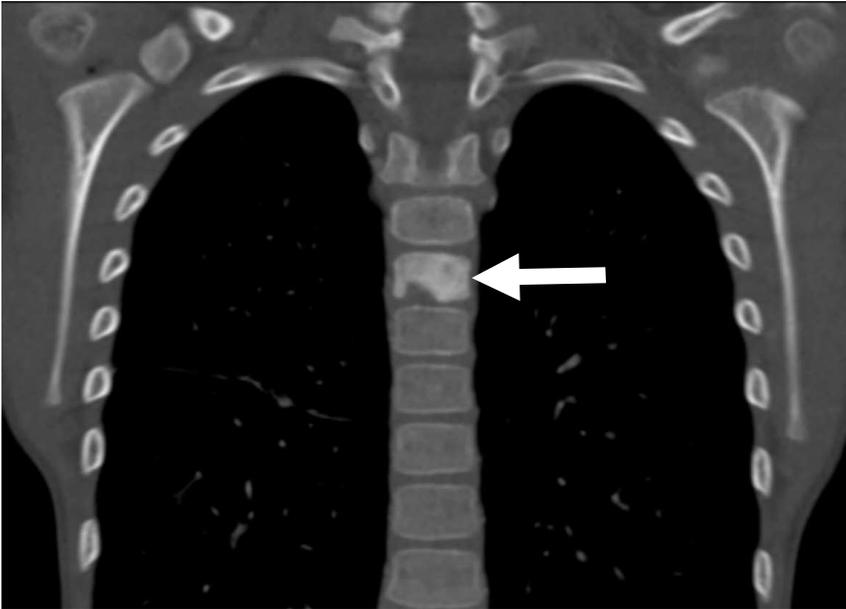
6 year old male presents for immunodeficiency evaluation for 6 weeks of nocturnal fevers, body aches, and fatigue

HPI: Began to have frequent “asthma exacerbations” and otitis media 6 months prior to visit

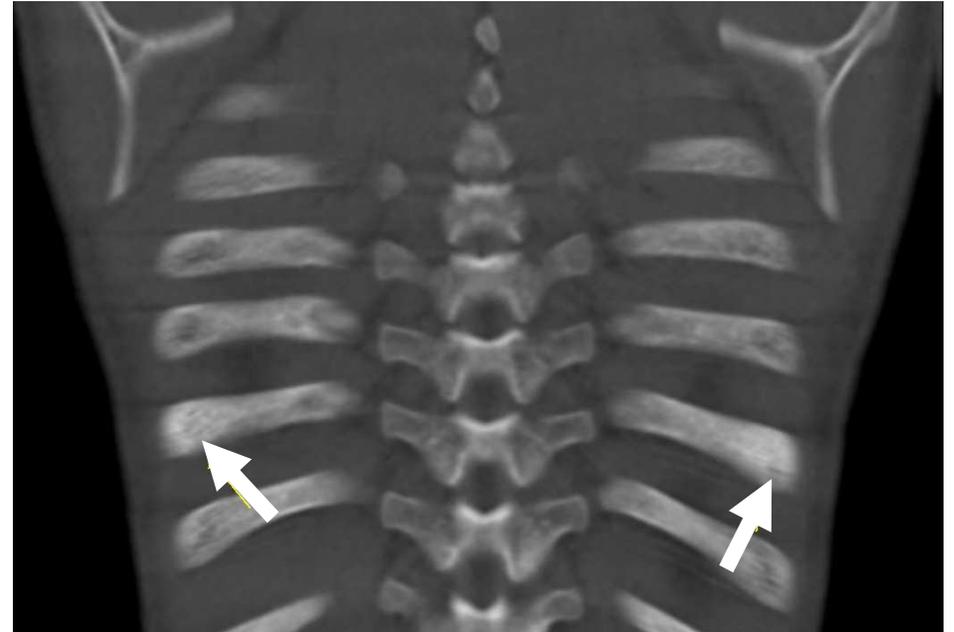
Past Medical History

- Born full-term to non-consanguineous family
- Diagnosed with asthma at the age of 6 months
- Admitted and treated for pulmonary *Mycobacterium avium* complex (MAC) infection at 2 years of age
 - Completed 2 years of treatment--azithromycin, rifampin, ethambutol, and ciprofloxacin
 - CT imaging of his chest 14 months into treatment showed a multi-cystic lesion in the right lung base

HRCT revealed sclerosis of bones



Vertebrae

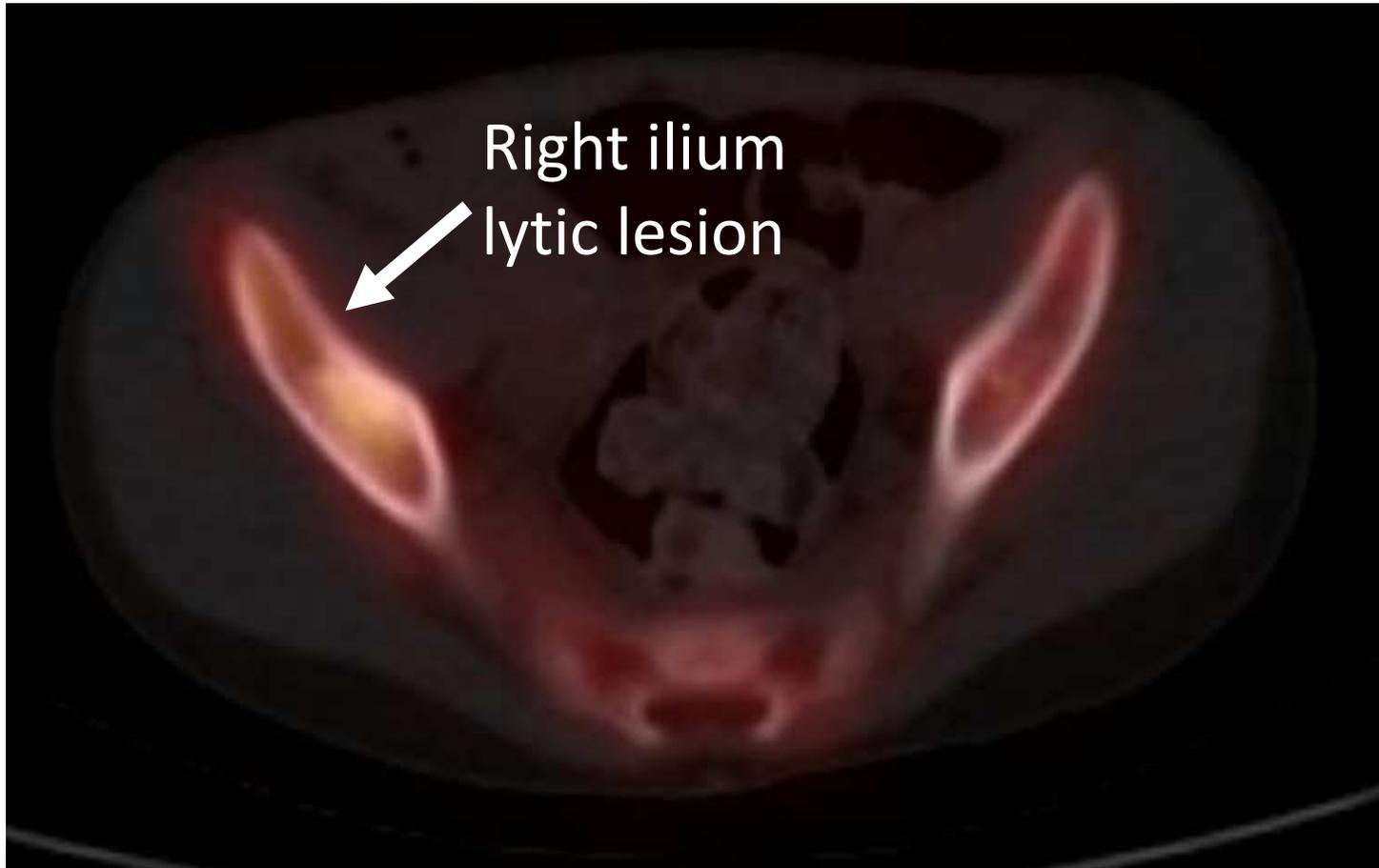


Ribs

Laboratory Results

Laboratory Test	Pertinent Results
CBC with Differential	↓ Hgb/Hct, ↑ platelets, ↑ monocytes
Inflammatory Markers	↑ CRP, ↑ ESR
Immunoglobulins	↑ IgG with ↑ IgG2 and IgG4
Antigenic T-cell Stimulation	↓ by 75% (spontaneous normal; response to candida and tetanus decreased)
Cytotoxic Lymphocyte Function	↓ cytotoxic lymphocyte function with absent CTL lytic units
Plasma cytokines	↑ IL-6
STAT1 phosphorylation	Normal
Other Tests Normal	Lymphocyte subpopulation, B cell number/maturation, vaccine responses, toll-like receptor function

Bone Scan/PET Scan



Disease Course

- “Back pain and fatigue” somewhat improved
- In vitro STAT1 phosphorylation in response to interferon-gamma and interferon-alpha normal
- Sequencing of the *STAT1* gene revealed a novel, heterozygous missense mutation, c.1378A>G (p.N460D), in the DNA-binding domain
- PET scan at 6 month F/U revealed new lytic lesions, but no changes in previous lesions
 - Biopsy of left acromion lesion grew MAC
 - ID: Ethambutol, Rifampin, and Azithromycin
- HLA typing for donor search for BMT: >50 full matched donors

Acknowledgements

- Zeynep Yesim Kucuk, MD (BMT-PID-CCHMC)
- Alan Brody, MD (Radiology-CCHMC)
- Elizabeth Sampaio MD, PhD (NIH-NIAID-LCID)
- Steve Holland MD, (NIH-NIAID-LCID)
- Sergio Rosenzweig, MD, PhD-PID Summer School Mentor

Extras

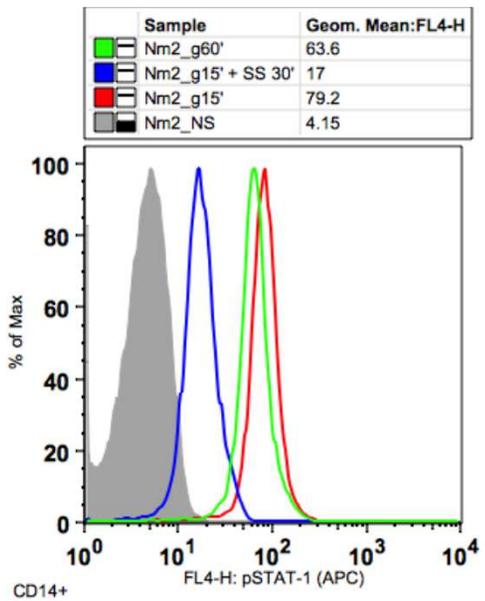
Next slides are for possible questions/further information

Questions arise:

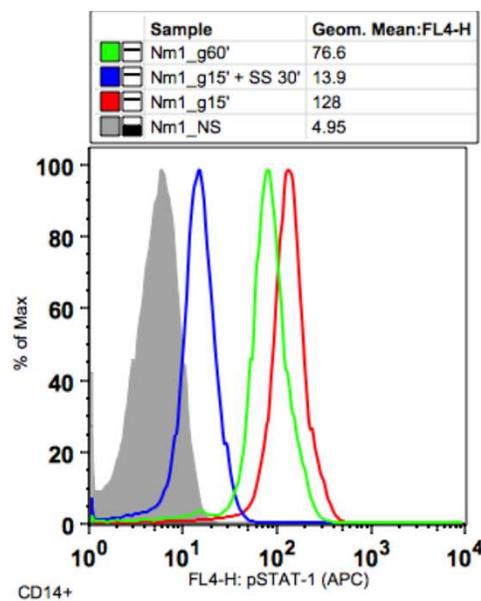
- Would you consider proceeding to HSCT? If so, when?
- Would you consider adding interferon-gamma onto treatment? Why or why not?

STAT1 Phosphorylation: Decreased in patient, but no defect in dephosphorylation

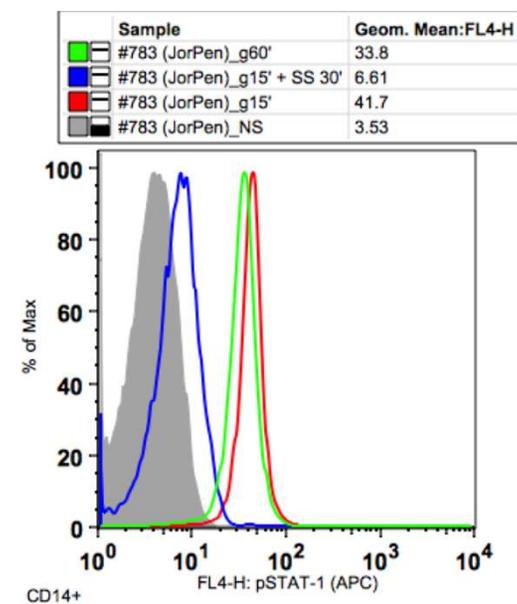
Normal 2



Normal 1

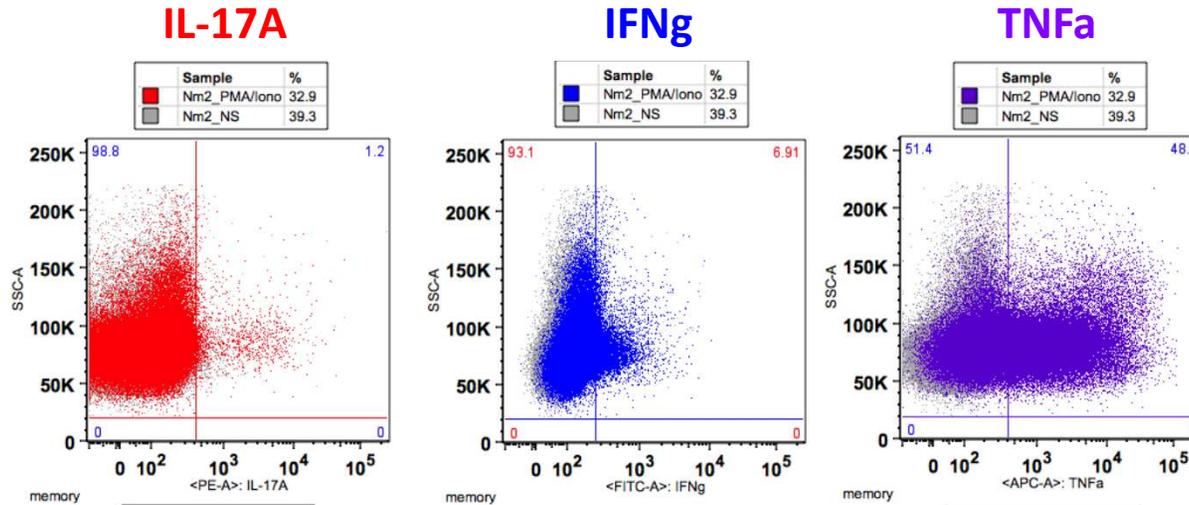


Patient

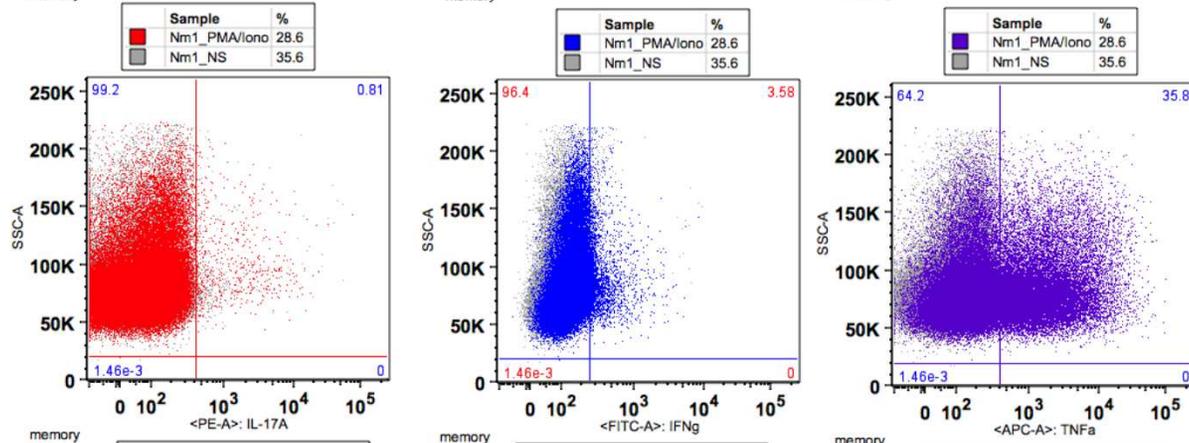


- Unstimulated
 - IFNg-400 for 15 min
 - IFNg-400 for 15 min, then staurosporine* for an additional 30 min
 - IFNg-400 for 60 min
- * (Tyrosine kinase-inhib.)

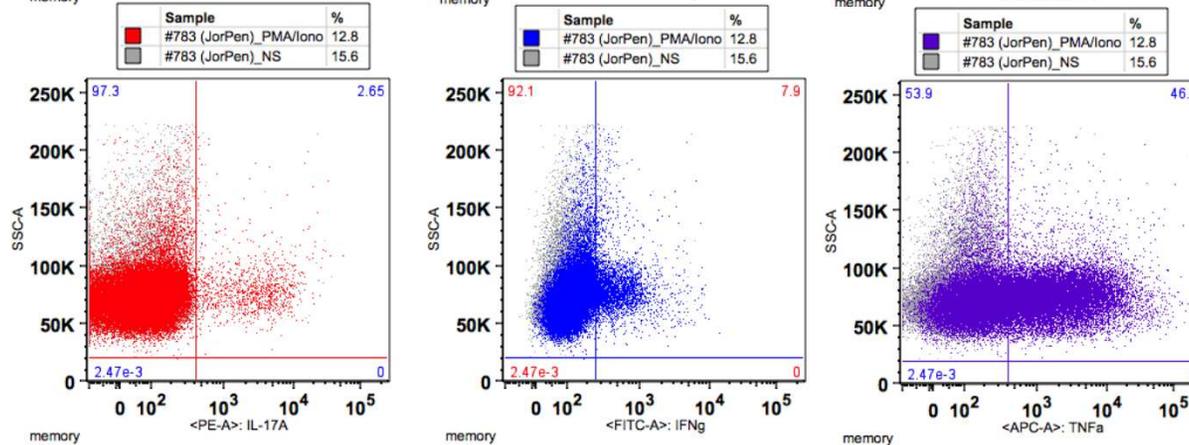
Normal 2



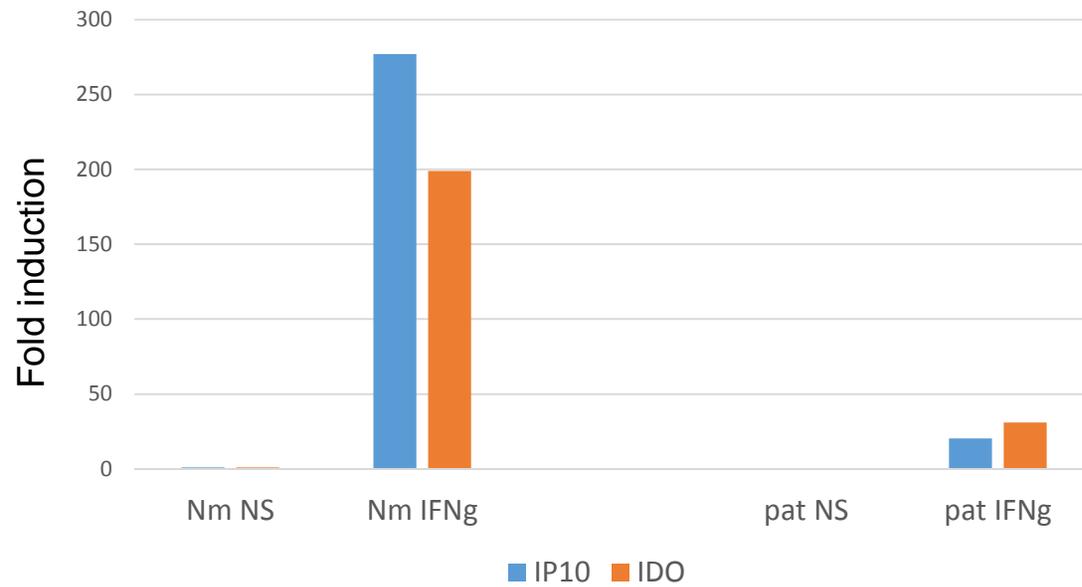
Normal 1



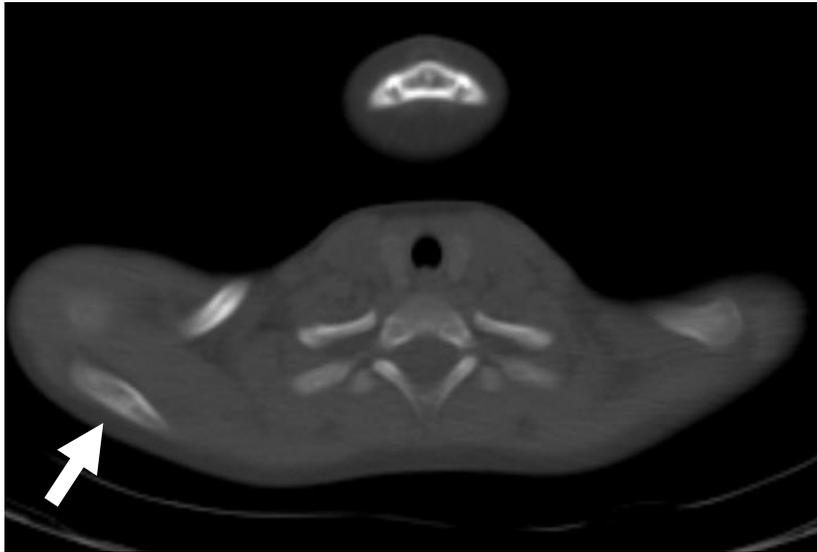
Patient



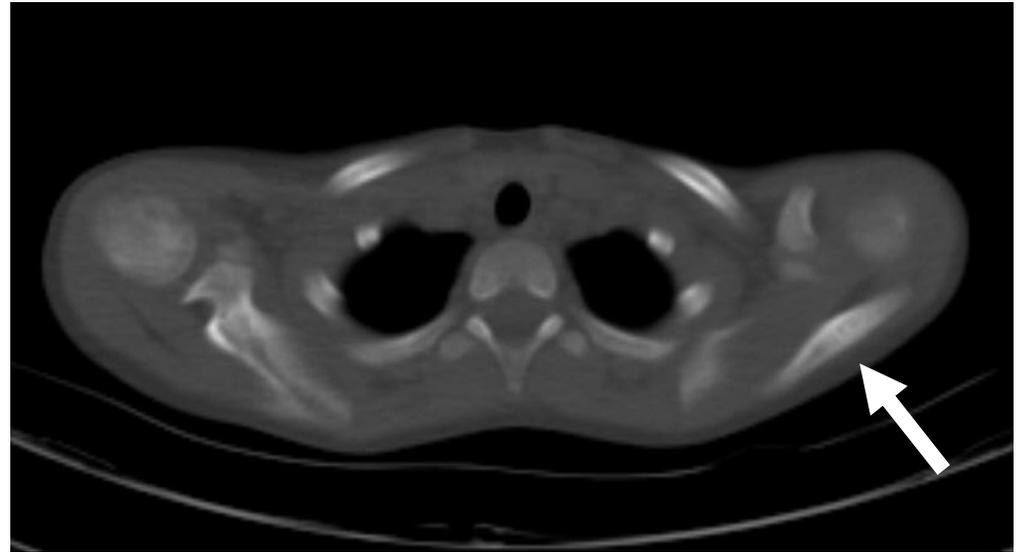
Decreased STAT1 Gene Expression



PET Scan

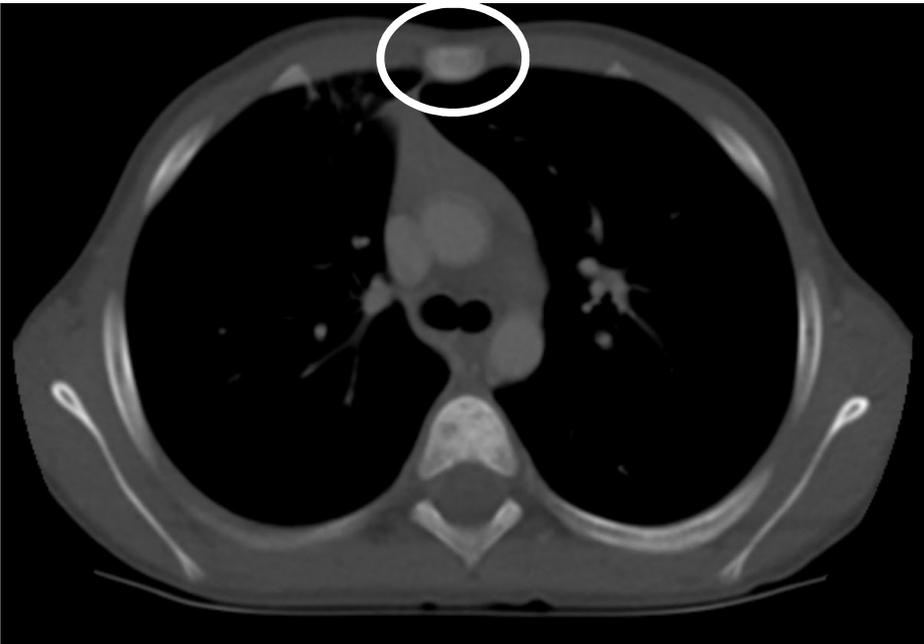


Normal Acromion



Abnormal Acromion

High Resolution CT Chest

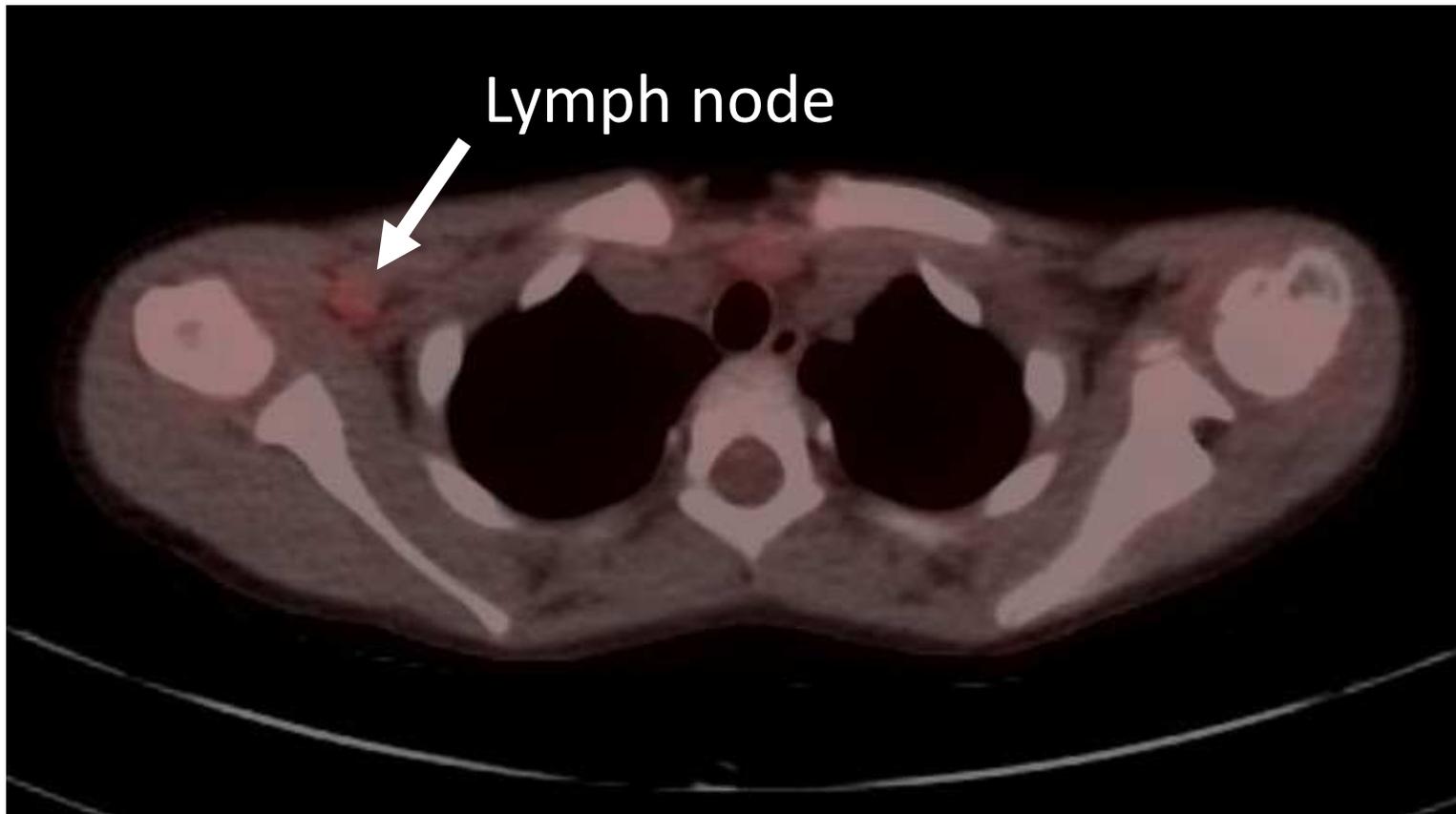


Normal Sternum



Abnormal Sternum

PET Scan



Laboratory Values

Laboratory Evaluation	Reference Ranges	Fall 2015	Summer 2016
WBC (K/mcL)	4.5-13.5K/mcL	12.8	8.7
Hgb (gm/dL)	12-16 gm/dL	11.4	12.5
PLT (K/mcL)	135-466K/mcL	762	498
Neutrophils	40-62%	53%	43%
Lymphocytes	15-45%	35%	41%
Monocytes	0-8%	8%	10%
Eosinophils	0-5%	2%	4%
Basophils	0-1%	1%	1%
ANC	1.8-8K/mcL	6.78	3.74
ALC	1.5-6.5K/mcL	4.61	3.57

Laboratory Evaluation	Reference Ranges	Fall 2015	Summer 2016
Auto Antibodies Screen		-	-
C3	71.0-150.0 mg/dL	209	159
C4	11.8-39.0 mg/dL	38.4	26.3
CH50	101-300	292	
C-Reactive Protein	<=0.30 mg/dL	5.64	0.40
Sedimentation Rate	0-20 mm/hr	84	27

Laboratory Evaluation	Reference Ranges	Fall 2015	Summer 2016
CD3	52-78% 800-3500 cells/mcL	71% 2824	71% 2757
CD4	25-48% 400-2100 cells/mcL	39% 1555	38% 1479
CD8	9-35% 200-1200 cells/mcL	24% 954	24% 952
CD19	(8-24%) 200-600 cells/mcL	22% 857	19% 752
CD16/56	6-27% 70-1200	6% 229	8% 307
CD4/CD8	1.0-3.6	1.6	1.6

Laboratory Evaluation	Reference Ranges	Fall 2015	Summer 2016
FOXP3+Tregs	55-81%	58%	
CD4+25+127 low Tregs	4.2-9.9% of CD4+	8.5%	
TCRV11/VA24	>=0.01%	0.06	
CD4+CCR6+CD45RA-*	10.7-27% of CD4+		13.3
CCR6+CD45RA-IL-17A+	1.2-4.6% of * cells	5.6	4.5
TCR-v-β		Normal expansion	
CD40L			
CD40L (Resting)	0-20%	0%	
ICOS (Resting)	0-29%	18%	
CD40L (Stimulated)	79-96%	71%	
ICOS (Stimulated)	74-97%	87%	

Laboratory Evaluation	Reference Ranges	Fall 2015	Summer 2016
Activation Markers			
CD69	0-3%	0%	
CD25	18-34%	22%	
CD71	0-25%	12%	
CD40L (CD154)	0-11%	1%	
CD134	0-12%	3%	
HLA-DR	0-15%	3%	
CD95	27-79%	34%	
Neutrophil Oxidative Burst		Normal Study	

Laboratory Evaluation	Reference Ranges	Fall 2015	Summer 2016
Plasma Cytokines			
IL-1B	<= 58 pg/mL	<10	<10
IL-2	<= 9 pg/mL	<5	<5
IL-4	<= 17 pg/mL	5	6
IL-5	<= 4 pg/mL	1	1
IL-6	<=7 pg/mL	39	<5
IL-8	<=47 pg/mL	4	6
IL-10	<= 7pg/mL	1	2
IL-18	89-540 pg/mL	82	94

Laboratory Evaluation	Reference Ranges	Fall 2015	Summer 2016
Intracellular Cytokines			
G-IFN			
CD4	8-24%	6%	4%
CD8	20-48%	16%	12%
NK Cells	48-80%	29%	5%
NKT Cells	18-64%	30%	NR (insufficient number of cells in sample)
TNF-a			
CD4	39-67%	18%	4%
CD8	17-61%	14%	6%
NK Cells	29-61%	18%	3%
NKT Cells	23-73%	30%	NR
IL-4			
CD4	<= 3.7%	0.2%	0.0%
CD8	<= 3.0%	0.0%	0.0%
NK Cells	<= 1.6%	0.1%	0.1%
NKT Cells	<= 3.1%	0.0%	NR

Laboratory Evaluation	Reference Ranges	Fall 2015	Summer 2016
IgG	560-1307 mg/dL	1370	1080
IgG1	400-1080 mg/dL	823	
IgG2	85-410 mg/dL	519	
IgG3	13-142 mg/dL	66	
IgG4	<=189 mg/dL	487	
IgM	60-263 mg/dL	153	127
IgA	68-378 mg/dL	191	116
IgE	2-307 IU/mL	137	
Mannan Binding Lectin	>= 50ng/mL	>3882	

Laboratory Evaluation	Reference Ranges	Fall 2015	Summer 2016
Auto Antibodies Screen		-	-
C3	71.0-150.0 mg/dL	209	159
C4	11.8-39.0 mg/dL	38.4	26.3
CH50	101-300	292	
C-Reactive Protein	<=0.30 mg/dL	5.64	0.40
Sedimentation Rate	0-20 mm/hr	84	27
Toll-Like Receptor Function Assay		Normal	
Vaccine Response		Normal	
Toll-Like Receptor Function Assay		Normal	

Laboratory Evaluation	Reference Ranges	Fall 2015	Summer 2016
B cell panel			
CD19+ B cells	10-31 %	20%	
CD19+CD27-CD21- IgM++	0.1-5.1 %	0.7%	
CD19+CD27-	64- 81 %	73%	
CD19+CD27-CD21+	60-74 %	69%	
CD19+CD27-IgM+IgD+	58-78 %	67%	
CD19+CD27-IgM+IgD+	91-98 % of CD27-	91%	
CD19+CD27-CD21- CD38-	0.5-2.9 %	1.0%	
CD19+CD27+	19-37 %	27%	
CD19+CD27+IgM+IgD+	8- 22 %	17%	
CD19+CD27+IgM+IgD+	42-72 % of CD27+	62%	
CD19+CD27+IgM+IgD-	0.7-5.2%	1.5%	
CD19+CD27+IgM+IgD-	2.8-16.0 % of CD27+	5.4%	
CD19+CD27+IgM-IgD-	3.5-14.4 %	7.9%	
CD19+CD27+IgM-IgD-	17-39 % of CD27+	29%	
CD38++CD138++	0.000- 0.032 % of Lymphocytes	0.020%	

Laboratory Evaluation	Reference Ranges	Fall 2015	Summer 2016
CD45RA/CD45R0			
CD3+	62.2-77%	71.7%	
CD4+ (of CD3)	39.8-69.4%	54.9%	
Naive	47.4-86.2%	70.3%	
Memory	13.5-52.0%	29.7%	
Naïve CD31+	69.1-95.6%	73.5%	
Naïve CD31-	4.2-30.7%	26.2%	
Memory Tcm	10.7-42.6%	27.0%	
Memory Tem	30.3-72.0	42.5%	
TEMRA	0.0-0.2%	0.0%	
CD8+ (of CD3)	21.6-44.3%	31.7%	
Naive	46.3-98.6%	67.8%	
Memory	1.1-42.4%	30.7%	
Memory Tcm	0.0-27.6%	3.6%	
Memory Tem	7.6-74.1%	44.9%	
TEMRA	0.0-12.7%	1.5%	

Laboratory Evaluation	Reference Ranges	Fall 2015	Summer 2016
NK Cell Function			
NK 50:1	>= 20%		12%
NK 25:1	>=10%		7%
NK 12:1	>=5%		4%
NK 6:1	>=1%		3%
NK Lytic Units	>=2.6		2.5
CD16/56% Positive	4-26%		10%
Cytotoxic T Lymphocyte Function			
CTL 50:1	>=35%	9%	
CTL 25:1	>=25%	5%	
CTL 12:1	>=15%	3%	
CTL 6:1	>=10%	2%	
Lytic Units	>=6.0	0	

Laboratory Evaluation	Reference Ranges	Fall 2015	Summer 2016
Lymphocyte Proliferation to Antigen Stimulation			
Spontaneous Response	0-280 cpm	853 cpm	4361 cpm
Phytohemagglutinin		Mitogen not available	197465 cpm
Candida albicans	>= 15289 cpm	2598 cpm	59442 cpm
Tetanus	>=4761 cpm	1085 cpm	61224 cpm
Quantiferon TB Gold		Negative	-
CMV IgG/IgM		- / -	
EBV IgG/IgM/EBNA		+ / - / +	